

Phosphorus efflux from maize roots is highly localised to the root tip

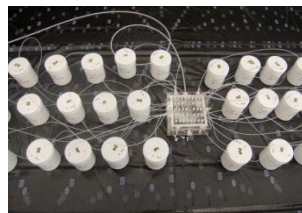
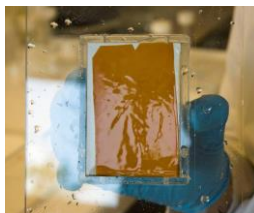
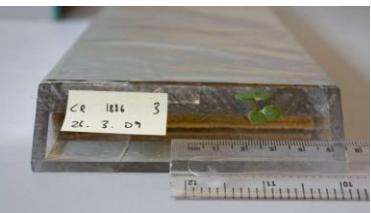


University of Natural Resources
and Life Sciences, Vienna

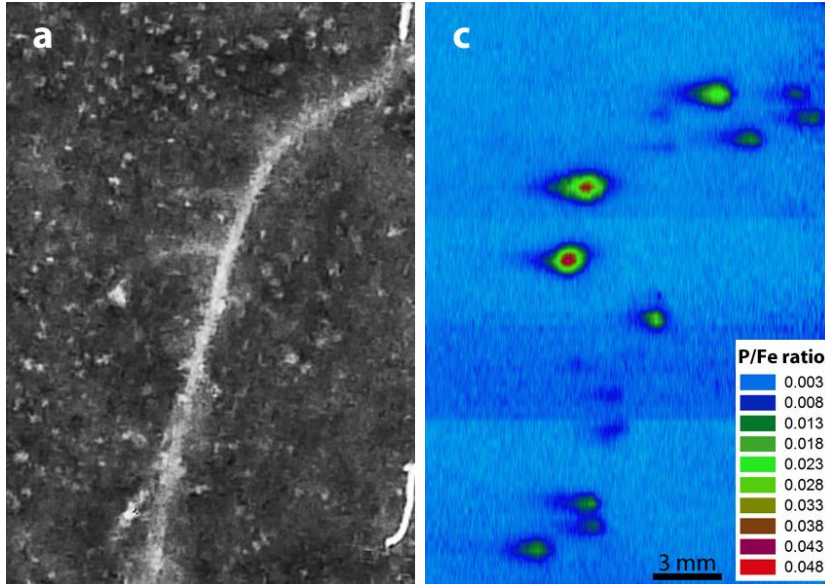
Jakob Santner, Rainer Muehlbacher, Andreas Kreuzeder and Walter W. Wenzel

Institute of Soil Research, University of Natural Resources and Life Sciences, Vienna, Austria

Thanks to **Philip White** (The James Hutton Institute) for data discussion.

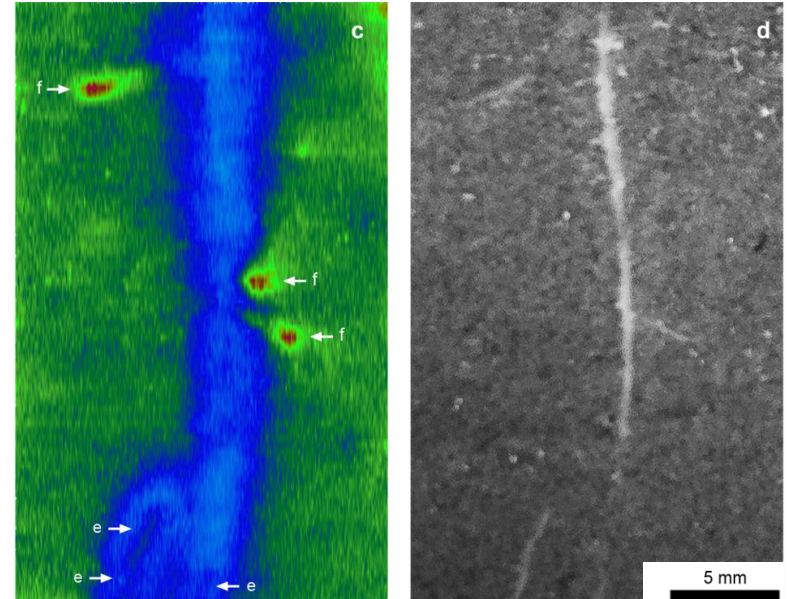


Introduction Phosphorus uptake & release in rapeseed and maize



Maize

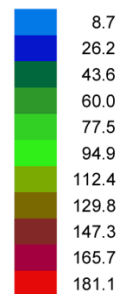
Santner et al., unpublished.



Rapeseed

Santner et al. 2012, Exp. Env. Bot.

$P C_{DGT}$ (nmol L⁻¹)



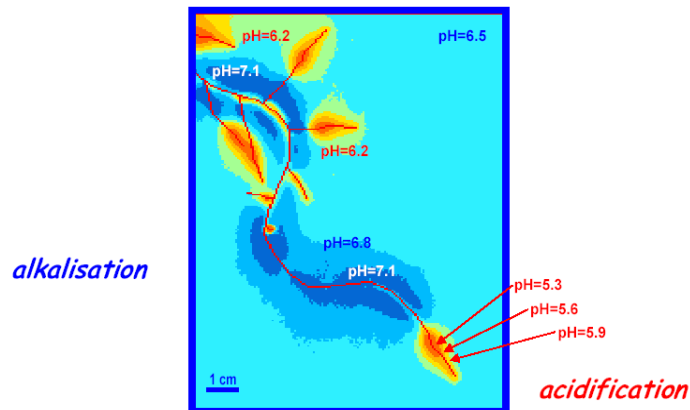
Introduction What causes the P hotspots?

P released from soil by root exudates (H^+ , carboxylates)?

- Desorption from soil?
- Dissolution of PO_4 minerals?

P released from directly from roots?

- Casparian strip permeable at root tip (Efflux)?
- Crushed root border cells?

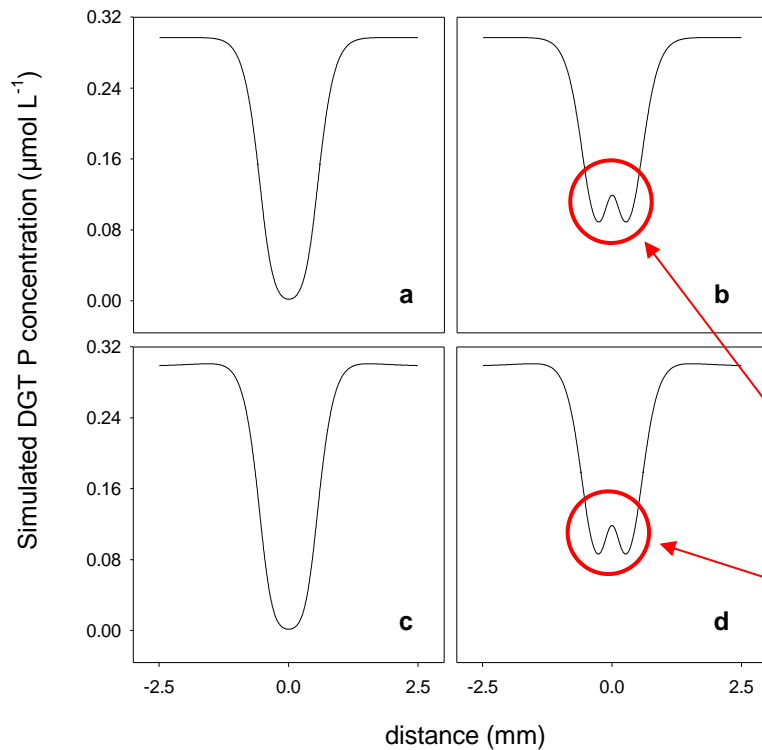


Ruiz 1992 – PhD thesis



Hinsinger et al. Plant & Soil 321, 2009

Introduction Numerical simulation points at efflux



Simulations:

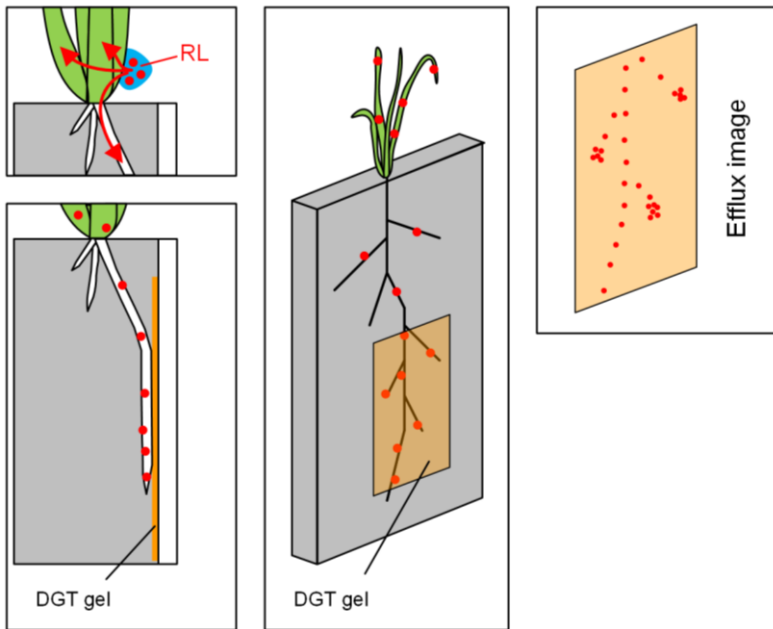
- a) No efflux, no citrate exudation
- b) Efflux only
- c) Citrate exudation only
- d) Efflux & citrate exudation

**Peaks caused by direct P release
(efflux)**

Santner et al. 2012, Exp. Env. Bot.

Question Do we see efflux?
Anything else?

Aim Image ^{33}P release from plant roots.



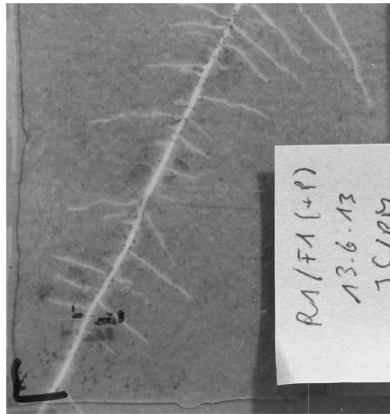
- (1) ^{33}P -containing droplet onto lesion (needle sting) on the coleoptile to radiolabel shoot P of maize.
- (2) ^{33}P is redistributed via the phloem.
- (3) Imaging of potential ^{33}P release from roots using the 'diffusive gradients in thin films' technique.

Results

Localisation of P efflux from maize roots

t_0

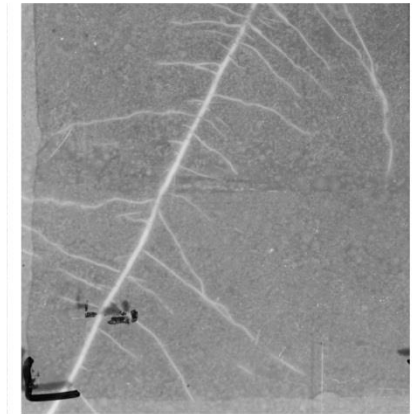
^{33}P label. & DGT app.



2 cm

t_{48h}

Note root growth!



Results

Localisation of P efflux from maize roots

t_0

^{33}P label. & DGT app.

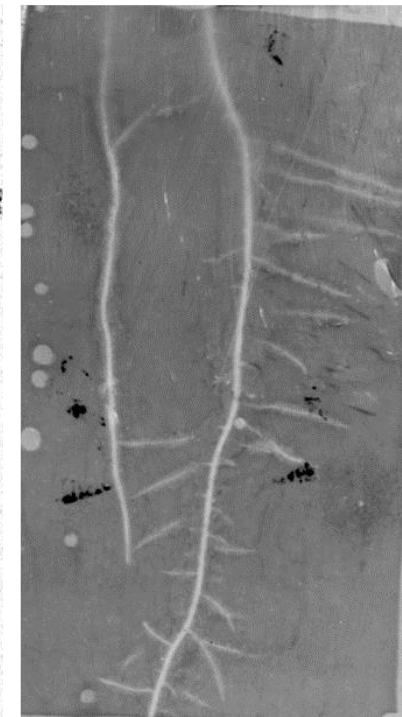
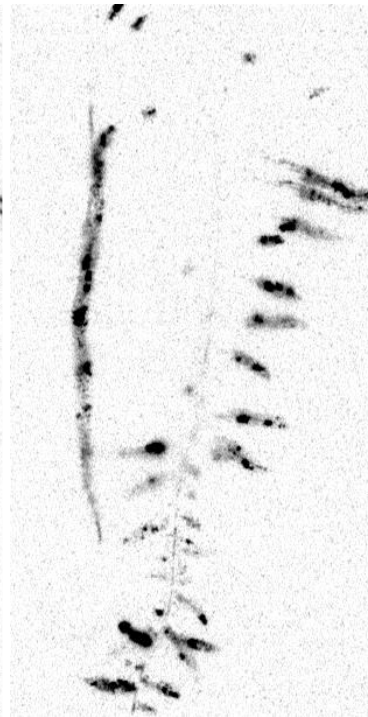
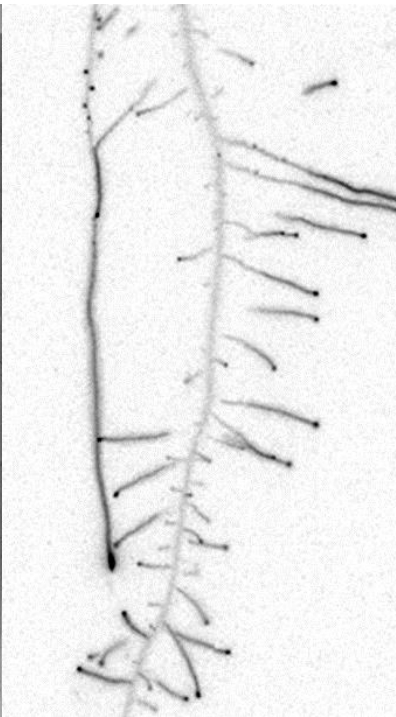
$t_{48\text{h}}$

total ^{33}P in the root

$t_0-t_{48\text{h}}$

^{33}P on the DGT gels

$t_{48\text{h}}$



5 cm

Results

Artefacts due to root injury?

Plant cultivation in agar medium to reduce risk of root injury.

t_0

^{33}P label. & DGT app.

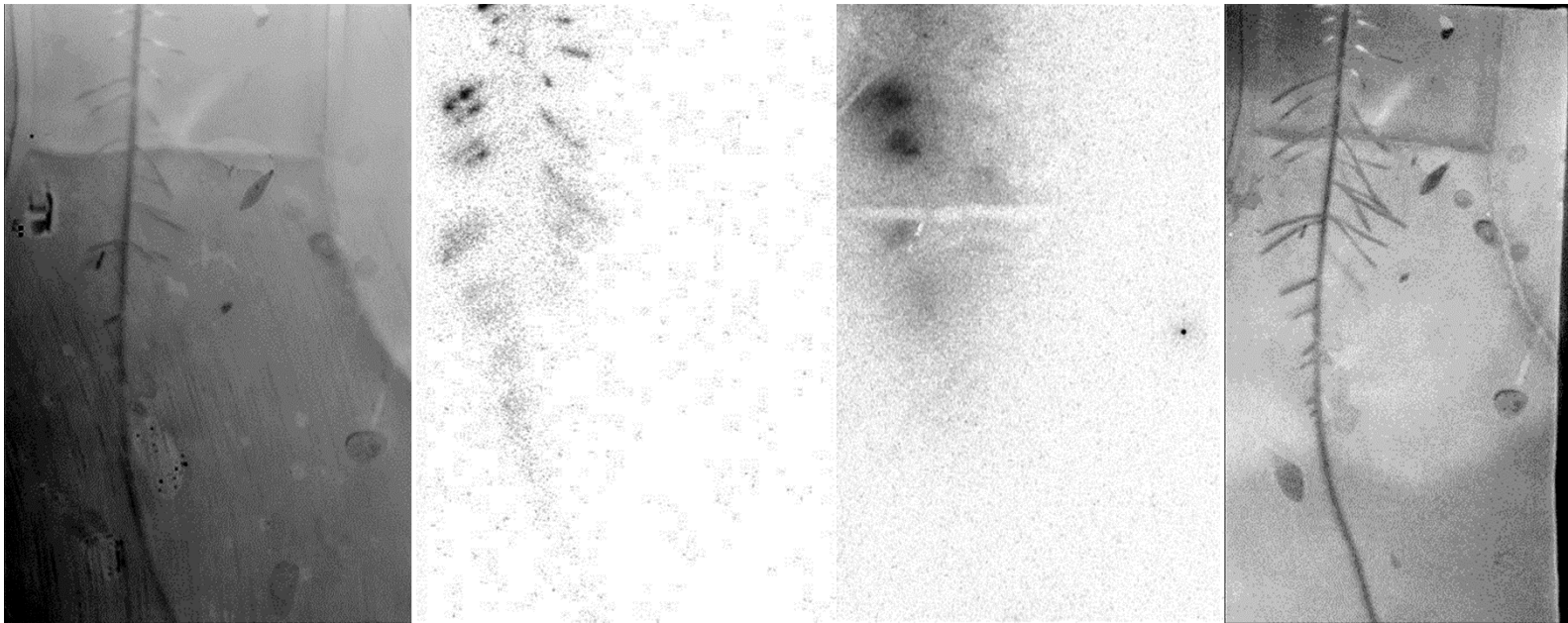
$t_{48\text{h}}$

total ^{33}P in the root

$t_0-t_{48\text{h}}$

^{33}P on the DGT gels

$t_{48\text{h}}$



5 cm

Results

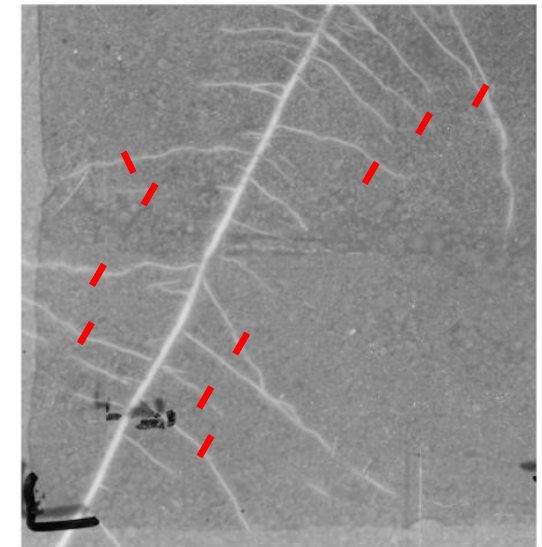
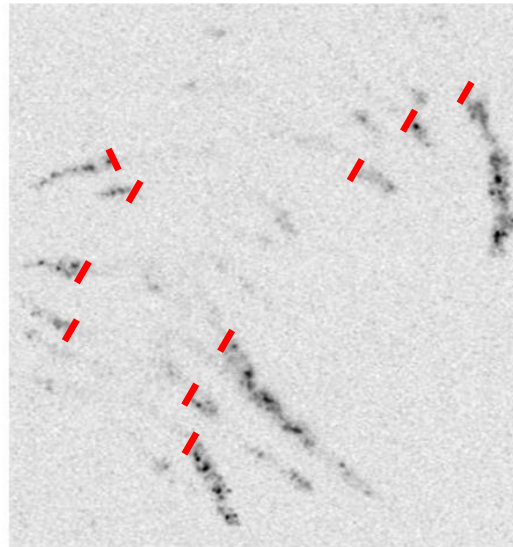
What's the size of the effluxing region?

Root growth: Efflux along newly grown area or confined to the very tip?

Start of Exp.



End of Exp. (48 h)



Almost all efflux from roots grown during sampling.

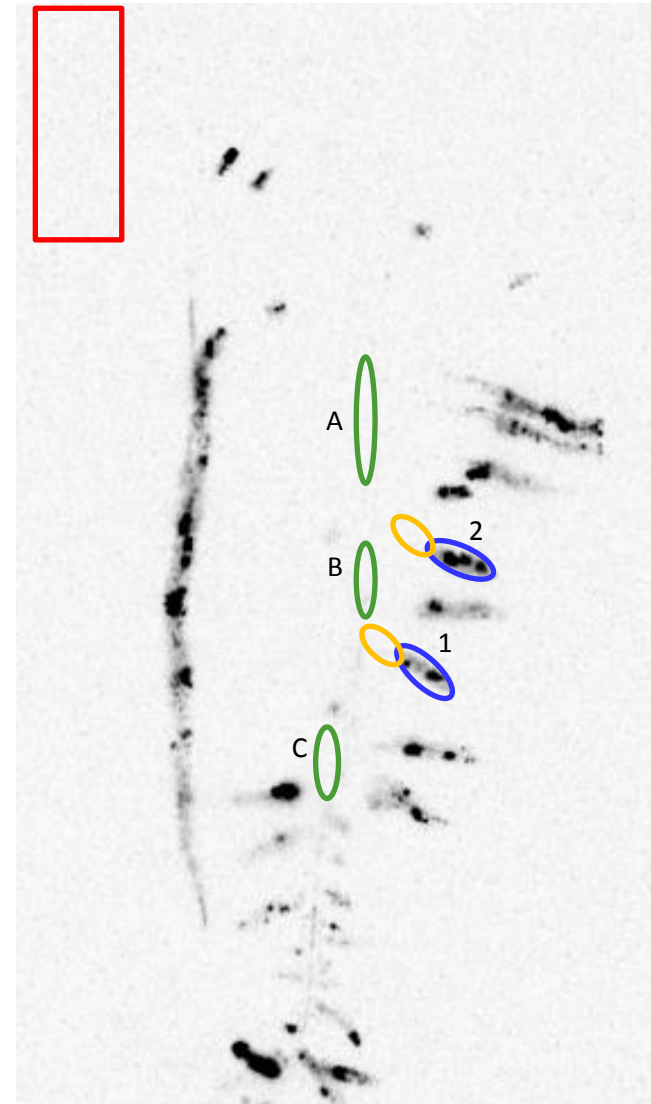
→ Indication for efflux from a very small area at the tip.

Measurement of ^{33}P efflux quantity in distinct image areas.

$$\frac{\text{efflux rate of root tip (cpm/cm}^2\text{)}}{\text{efflux rate of root axis (cpm/cm}^2\text{)}} \approx 300$$

Assumption: very small (1 mm^2) effluxing root tip area

Virtually all P is effluxed at the tip areas!



- Efflux was assumed to be homogeneously distributed across the root surface.

Classens & Barber 1974 Plant Phys.

$$I = \frac{I_{\max} c}{K_m + c} - E$$

$$I = \frac{I_{\max} (c - c_{\min})}{K_m + (c - c_{\min})}$$

- P uptake (influx) along major parts of the root axis is well documented.

Rubio et al. 2004 J. Exp. Bot., Rovira & Bowen 1968 Nature, ...

- **BUT:** Efflux cannot be subtracted from influx if both processes are not co-localised.

- c_{\min} much lower than assumed along the majority of absorbing root surfaces.
- Earlier work showed overestimation of K_m values due to diffusion-limited supply to the sites of apoplastic uptake.
(Santner et al. 2012, PCE; Degryse et al, 2012, Plant Phys.)

Further questions:

- Roots practically zero-sinks for P?
- Are MM kinetics unnecessary?
- Is it more important to accurately model soil P supply for modelling plant P uptake?

**Thanks for your
attention**